

Synthesis, characterization and antinociceptive activities of Novel 2-(2,4-dichlorophenyl)-4H-chromen-4-one

Mohammad Shoaib*¹, Syed Wadood Ali Shah¹, Shafiullah Shah¹,
Nawaz Khan¹ and Muhammad Naeem Ahmed²

¹Department of Pharmacy, University of Malakand, Khyber Pakhtunkhwa, Pakistan

²Department of Chemistry, University of Azad Jammu and Kashmir, Muzaffarabad, Pakistan

Abstract: A novel flavone derivative has been synthesized in good yield from ketone and aldehyde. The structure has been established by different spectroscopic techniques like ¹H NMR, ¹³C NMR, IR and elemental analysis. The compound was then screened for its acute toxicity and antinociceptive activity studies on animal model. The novel compound was safe upto a maximum dose of 500mg/kg body weight oral dose in mice and showed 65.92 and 82.18% peripheral analgesic activity at 15 and 30mg/kg body weight doses. Central antinociceptive activity of the compound was 53.13 and 64.44% at 15 and 30mg/kg body weight respectively.

Keywords: Flavonoids, chalcone, flavone, acute toxicity, antinociceptive activity.

INTRODUCTION

Flavonoids are natural products and important part of human diet that are abundantly present in vegetables, fruits, nuts, seeds, wine, tea and flowers. Flavonoids are bioactive polyphenols of low molecular weight that play a key role in photosynthesizing cells of plants (Sandhar *et al.*, 2011). On the basis of chemical structures, flavonoids are mainly classified into six classes, i.e., flavones, flavanones, flavans (flavanols), isoflavonoids, anthocyanins and flavonols. All these groups mainly differ by the structure of the C-ring and functional groups attached to C-3 and C-4 of the main structure (Peterson and Dwyer, 1998). Pharmacological role of flavonoids in human is a subject of extensive research and they have been known for a longtime to possess numerous biological activities both in animals and humans (Fernández *et al.*, 2006; Maikai, 2011).

Flavonoids exhibit several biological and pharmacological effects such as antihepatotoxic, anti-ulcer actions (Bors *et al.*, 1990; Smith *et al.*, 1988), strongradical scavenging and antioxidant activity (Zhang *et al.*, 2006; Chan *et al.*, 2000; Rice-Evans *et al.*, 1996). They also appears to be associated with reduction of certain chronic ailments (Kris-Etherton *et al.*, 2004), like cardiovascular complications (Gross, 2004) and various kind of cancers (Moon *et al.*, 2006). Flavonoids exhibit antimicrobial (Cushnie and Lamb, 2005), antiviral (Asres *et al.*, 2005), antiulcer (Wightman, 2004), anti-inflammatory activities (Kim *et al.*, 2004, Dao *et al.*, 2004), anti-allergenic (Middleton and Kandaswami, 1992; Berghe *et al.*, 1993), beneficial effects on capillary fragility (Benavente-García *et al.*, 1997), an ability to inhibit human platelet aggregation (Tijburg *et al.*, 1997),

antipyretic (Ray *et al.*, 2006), analgesic (Ahmed *et al.*, 2007) and antidepressant (Fan *et al.*, 2012) properties.

Chalcones are an important class of compounds which are good intermediates for the synthesis of various heterocyclic compounds like flavones, flavanols, flavanones, isoxazolines and aurones (Shah and Goswami, 2013). Different methods have been reported for flavone synthesis like Allan-Robinson synthesis, synthesis from chalcones, Wettig method, from o-benzoyl acetophenone and synthesis under catalytic microwave irradiation (Mostahar *et al.*, 2006). These flavones have antioxidant (Gabrielska *et al.*, 1997), antimicrobial (Husain *et al.*, 2013), anticancer (Zhang *et al.*, 2008), anti-inflammatory (Carvalho *et al.*, 1999), analgesic (Thirugnanasambantham *et al.*, 1993) and antidepressant (Fan *et al.*, 2012) activities.

Based on the reported literature regarding the synthesis of flavones from chalcones and their potentials to have biological and pharmacological activities, we have synthesized and explored antinociceptive effects of a novel flavone on scientific grounds.

MATERIALS AND METHODS

General

Chemicals/ reagents used for the synthesis of the flavone were of Sigma Chemical Company (Aldrich, Germany). Solvents like ethanol, *n*-hexane, ethyl acetate etc were synthesis grade purchased from E. Merck and used as such without prior distillation.

¹H-NMR and ¹³C NMR spectra were recorded in deuterated chloroform (CDCl₃) on Bruker SF spectrometers operating at 300 and 75 megahertz (MHz) frequencies, respectively. Chemical shifts values are

*Corresponding author: e-mail: pharmacistsyed@gmail.com

expressed in δ (ppm) downfield relative to TMS which was used as an internal standard. Infrared spectra were recorded on Thermo scientific USA (Nicolet 6700), Infrared spectrometer on KBr disk method. All melting points are uncorrected and were taken in open capillary tubes using Electrothermal 9100 apparatus (Barnstead UK). Reaction extents and final products purities were checked on TLC plates (Merck 60 F₂₅₄, Darmstadt Germany) and spots were visualized under UV Lamp (180-365 nm) and with subsequent staining with iodine vapours.

Synthesis of 2-(2,4-dichlorophenyl)-4H-chromen-4-one derivative (F5)

1.80mL of 2-hydroxy acetophenone (15 mmol) was added to absolute ethanol (20mL) in a round bottom flask and then sodium hydroxide solution 10mL (40% ethanolic) was added drop wise to this solution at room temperature. Then 2.62gm of 2,4-dichlorobenzaldehyde (15mmol) in ethanol was added dropwise to this mixture and stirred for 24 hours at room temperature (25±2°C). The reaction was monitored by TLC using ethyl acetate: *n*-hexane (2:8 v/v). Upon completion of the reaction, it was poured into crushed ice and neutralized with 1N HCl solution resulting in yellow precipitates of chalcone. The chalcone was filtered and washed with water to remove the impurities.

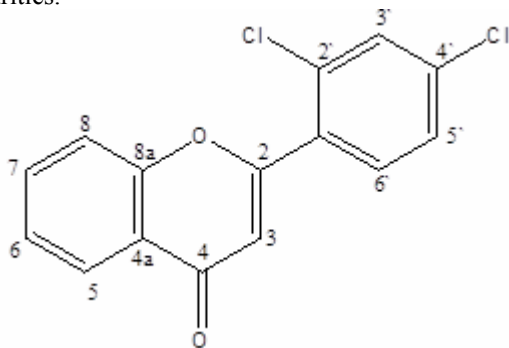


Fig. 1: Structure of the synthesized flavone F5 (2-(2,4-dichlorophenyl)-4H-chromen-4-one)

In next step, chalcone was cyclized to flavone in 15mL DMSO in the presence of iodine (375mg) at 140°C for 1 hour and the extent of reaction was monitored by TLC. Upon completion of reaction, mixture was cooled to room temperature and was poured into water followed by extraction with ethyl acetate (25mL×3), treated with sodium thiosulphate solution (20%), brine solution and dried over sodium sulphate. The final product (mixture of flavones and chalcone) was subjected to gravity column chromatography. The flavone (fig. 1) was obtained as white solid on elution with *n*-hexane: ethyl acetate (9:1).

Pharmacological activities

Drugs and animals

Tween-80 was purchased from Daejung Chemicals, Korea. Diclofenac sodium and Morphine sulphate were purchased from local market. Swiss Albino mice of either

sex weighing 30-35g were purchased from National Institute of Health (NIH) Islamabad. The animals were housed in individual cages at the animal house of University of Malakand with free access to water and standard diet and starved for 12-18 hours before experimentation. Ethical Committee of the Department of Pharmacy, University of Malakand approved the experimental protocols and ensured its compliance with provisions of the “Animal Bye-Laws 2008, Scientific Procedures Issue-I of the University of Malakand”.

Acute toxicity

The synthesized compound (F5) was subjected to acute toxicity study on mice. In first phase, three groups of overnight fasted mice (n=6) were given an oral dose of 50, 75 and 150 mg/kg body weight of synthesized novel flavone. In second phase, further 3 groups were given an oral dose of 300, 400 and 500 mg/kg body weight. All the treated animals were kept under observation for 24 hours. Finally, LD₅₀ was calculated in all groups by recording the number of deaths in each group within 24 hours (Lorke, 1983; Ali *et al.*, 2011).

Acetic acid-induced writhing test (peripheral activity)

Antinociceptive response of the novel synthesized compound (F5) was assessed by using mice as an animal model with slight modifications (Koster *et al.*, 1959). The synthesized compound at different dose concentrations in 2% tween-80 and 2% tween-80 in normal saline were dispensed to respective groups of animals (n=6) orally, and Diclofenac sodium was administered intraperitoneally. After 30 minutes of time interval, 0.1 mL of acetic acid at a concentration of 1% (v/v) was injected intraperitoneally. Data was recorded in the form of number of writhes (constriction of abdomen, turning of trunk and extension of hind legs) during 30 minutes of test period, beginning 3 minutes after the injection of acetic acid (Razmi *et al.*, 2013).

Tail immersion test (central activity)

Albino mice were used to assess the central antinociceptive response of the synthesized novel compound. Briefly, different dose concentrations of the synthesized compound and vehicle (2% tween-80) were administered orally 30 minutes and morphine (standard) intraperitoneally 15 minutes before the screening to respective groups. The latency period (time taken by the mice to deflect the tail) was determined by immersing 1-2 cm of mice tail in water that was kept warm at 53±1°C (Imam and Sumi, 2014).

STATISTICAL ANALYSIS

Data are presented as mean ±SEM. Analysis of variance and Dunnett's test is statistically manipulated with Graph Pad In Stat Version 3.10 software.

Table 1: Acetic acid induced analgesic activity data of the compound

Treatment	Dose (mg/kg)	Number of writhing	% inhibition
Control	2 % w/v of Tween 80	74.83±1.01	-----
Standard	Diclofenac sodium 100 mg	10.16±0.70**	86.42
F5	15 mg	25.50±0.84**	65.92
	30 mg	13.33±0.80**	82.18

All the values were expressed as mean ±SEM (n=6). **P<0.01 when compared to control group.

Table 2: Percent analgesic activity (Tail flick) data of the compound

Treatment	Dose	Time in Sec/% response					
		15min	30min	45min	60min	75min	90min
Control	2% w/v of Tween 80	0.628±0.04	0.698±0.01	0.686±0.02	0.633±0.01	0.623±0.01	0.641±0.01
Standard	Morphine sulphate 5mg	0.77±0.01*** (18.92)	1.13±0.04*** (38.38)	1.61±0.021*** (57.52)	2.241±0.02*** (71.74)	3.53±0.03*** (82.35)	2.33±0.05*** (72.50)
F5	15mg	0.685±0.07* (8.27)	0.79±0.06* (11.6)	0.958±0.02*** (28.34)	1.041±0.02*** (39.2)	1.33±0.01*** (53.13)	1.21±0.25* (47.26)
	30mg	0.73±0.01** (14.31)	0.928±0.10** (24.77)	1.05±0.04*** (34.70)	1.316±0.02*** (51.89)	1.75±0.02*** (64.44)	1.325±0.03*** (51.57)

All the values were expressed as mean ±SEM. *P<0.05, **P<0.01 and ***P<0.001 when compared to control group.

RESULTS

Chemistry

Proton NMR of F5 shows singlet at δ 6.68ppm referring to the methene proton, ¹³C NMR of this compound show fifteen different peaks at respective chemical shifts. Signal at δ 177.9 ppm show characteristic peak for ketonic carbon in ¹³C NMR. IR spectrum of the compound show peaks at 3066.5, 2920.6, 1734.1, 1645.4, 1221.1 and 748.2cm⁻¹ which confirms the presence of C-H Aromatic, C-H, methene, C=C, C=O, C-O and C-Cl bonds. CHN data calculated for compound C₁₅H₈Cl₂O₂ is: C, 61.88; H, 2.77. and Found is: C, 60.99; H, 2.28. This confirms the synthesis of the novel flavone.

Yield: 87%, m.p: 90.5°C, R_f: 0.57 (ethyl acetate: n-hexane (3:7)), ¹H NMR (300 MHz, Chloroform-d, ppm) δ 8.27 (dd, J=8.0, 1.7 Hz, 1H, H-3'), 7.74 (ddd, J=8.7, 7.1, 1.7 Hz, 1H, 5'-H), 7.64-7.56 (m, 2H, 5-H, 6'-H), 7.55-7.40 (m, 3H, 6-H, 7-H, 8-H), 6.68 (s, 1H, H-3). ¹³C NMR (75 MHz, CDCl₃, ppm) δ 177.98 (C-4), 161.51 (C-2), 156.54 (C-8a), 137.43(C-2'), 134.06 (C-4'), 133.81 (C-6'), 131.42 (C-5'), 130.77 (C-3'), 130.40 (C-1'), 127.58 (C-7), 125.82 (C-6), 125.50 (C-5), 123.81 (C-4a), 118.18 (C-8), 113.16(C-3). IR (KBr) V_{max}cm⁻¹: 3066.5, 2920.6, 1734.1, 1645.4, 1221.1, 748.2. Anal. Calcd. For C₁₅H₈Cl₂O₂: C, 61.88; H, 2.77. Found: C, 60.99; H, 2.28.

Acute toxicity

In our study, we evaluated the acute toxicity study in mice model. The animals were given different doses of the synthesized compound in different stages of the study. To a highest dose of 500mg/kg body weight, the compound did not show any mortality.

Analgesic activity (peripheral activity)

In our study for evaluation of peripheral analgesic activity of the synthesized flavone, we used the acetic acid induced mice model. Results of the activity are summarized in table 1. From the results it is evident that the synthesized compound showed 65.92 and 82.18% analgesic activity at 15 and 30mg/kg body weight, respectively in comparison with standard diclofenac sodium i.e 86.42%. From table 1, it is clear that the flavone at 30mg showed almost the same activity like that of standard.

Tail immersion test (central activity)

The latency time of mice to thermal stimuli in this study induced by hot water was significant in both doses at 15 and 30mg/kg body weight however the time for 50% of analgesic response in both doses is different (table 2). At 15mg/kg body weight, the compound showed 50% response in 75 minute while it was observed in 60 minute for 30mg/kg body weight. The standard drug morphine showed 50% response at 45 minute. Both doses showed highly significant activity at 45-75 minutes (P<0.001). Maximum effect was recorded for both doses of the compound at 75min. Based upon above findings, it may be assumed that the novel flavone possess good peripheral and central antinociceptive activities.

DISCUSSION

To evaluate pharmacological screening of a compound whether from plant or synthetic sources, its acute toxicity study should be carried out first. Acute toxicity screening in animal model is important preliminary test that provides basis for the dose estimation and predicts

potential toxicity of the drug. From this study, nature of acute response in humans can be anticipated and also provide rough idea about of the organ system involvement (Bhardwaj *et al.*, 2009). To a highest dose of 500 mg/kg body weight, the compound did not show any mortality. So up to this dose the compound is safe (Bala *et al.*, 2013).

Analgesic activity (peripheral activity)

Analgesic drugs alleviate pain symptoms without terminating its etiologic cause (Tripathi, 2013). Available analgesics classes like NSAIDs and opiates has adverse effects and are not considered useful in all respects. For this purpose, to improve the pain relieving quality of analgesics and to produce analgesics with less side effects, new agents are constantly sought out (Akter *et al.*, (2008). Writhing induced by acetic acid assay is a rapid and sensitive way of measuring peripheral analgesic potentials of compounds. Mechanism of the writhing induction is increased concentration of PGE2 and PGF2 produced by acetic acid (Collier *et al.*, 1968; Bentley *et al.*, 1983). For evaluating central analgesic effect of compounds, tail flick or hot plate methods are adopted. These methods were for the first time successfully described by Woolfe and MacDonald (Woolfe and MacDonald, 1944). Sensory nerves sensitize nociceptors and the involvement of substances like prostaglandins (PGs) are minimized in this activity (Mustaffa *et al.*, 2010). Analgesia of spinal origin and pain sensation to thermal stimulus is well demonstrated by the tail immersion in hot water and hot plate studies using mice for various opioid derivatives and other drugs (Subhan *et al.*, 2008).

As discussed earlier that acetic acid increases the concentration of prostaglandins (PGs) that cause the peripheral pain response (Bentley *et al.*, 1983) and NSAIDs like diclofenac inhibits the productions of PGs (Simmons *et al.*, 2004). So it is assumed that the analgesic activity of the flavone might be due to such mechanisms like that of the standard.

Tail immersion test (central activity)

Response of tail withdrawal in mice is considered to be centrally mediated (Srinivasan *et al.*, 2003). The latency time of mice to thermal stimulus is measured in both tail flick and hot plate tests. Hot plate method measures supraspinal reflexes mediated through μ 1- and μ 2-opioid receptors while tail immersion measures spinal reflexes of μ 2- and δ -opioid receptors (Imam and Sumi, 2014). Based upon above findings, it may be assumed that the novel flavone possess good peripheral and central antinociceptive activities.

CONCLUSION

Results of this study indicates that the synthesized compound possess both central and peripheral analgesic

activities. The peripheral activity is more significant than the central analgesic activity. Further elucidation of the compound for evaluation of its exact mechanisms of analgesic response is necessary.

REFERENCES

- Ahmed A, Ilyas NH, Ibrahim, Musa KY and Yaro AH (2007). Analgesic effects of *Tacazzea apiculata* Oliv. *Nigerian. J. Pharma. Sci.*, **6**(2): 134-138.
- Akter R, Hasan SR, Siddiqua SA, Majumder MM, Hossain MM, Alam MA, Haque S and Ghani A (2008). Evaluation of Analgesic and Antioxidant Potential of the Leaves of *Curcuma alismatifolia* Gagnep. *Stamford. J. Pharma. Sci.*, **1**: 3-9.
- Alcaraz L, Blanc OS, Puig O, Tomas F and Ferretti F (2000). Antibacterial Activity of Flavonoids Against Methicillin-resistant *Staphylococcus aureus* strains. *J. Theoretical Biol.*, **205**: 231-240.
- Ali N, Ahmed G, Shah SWA, Shah I, Ghias M and Khan I (2011). Acute toxicity, brine shrimp cytotoxicity and relaxant activity of fruits of callistemon citrinus curtis. *BMC Complemen. Altern. Med.*, **11**: 99.
- Asres K, Seyoum A, Veeresham C, Bucar F and Gibbons S (2005). Naturally derived anti-HIV agents. *Phytotherapy Res.*, **19**: 557-581.
- Bala S, Kamboj S, Saini V and Prasad D (2013). Anti-Inflammatory, Analgesic Evaluation and Molecular Docking Studies of N-Phenyl Anthranilic Acid-Based 1, 3, 4-Oxadiazole Analogues. *J. Chem.*, 2013.
- Benavente-García O, Castillo J, Marin FR, Ortuño A and Del Río JA (1997). Uses and properties of citrus flavonoids. *J. Agri. Food Chem.*, **45**: 4505-4515.
- Bentley G, Newton S and Starr J (1983). Studies on the antinociceptive action of α -agonist drugs and their interactions with opioid mechanisms. *Br. J. Pharmacol.*, **79**: 125-134.
- Berghe DV, Haemers A and Vlieunek A (1993). Bioactive natural products, detection and structural determination. CRC Press, London.
- Bhardwaj P, Kantharia N, Yadav P and Panwar A (2009). Acute toxicity study of an aqueous extract of *Ficus racemosa* Linn. bark in albino mice. *Internet J. Toxicol.*, **6**.
- Bors W, Heller W, Michel C and Saran M (1990). Flavonoids as antioxidants: determination of radical-scavenging efficiencies. *Methods. enzymol.*, **186**: 343.
- Chan EC, Pannangpetch P and Woodman OL (2000). Relaxation to flavones and flavonols in rat isolated thoracic aorta: mechanism of action and structure-activity relationships. *J. Cardiovasc. Pharmacol.*, **35**: 326-333.
- Collier H, Dinneen L, Johnson, CA and Schneider C (1968). The abdominal constriction response and its suppression by analgesic drugs in the mouse. *Br. J. Pharmacol. Chemother.*, **32**: 295-310.

- Cushnie T and Lamb AJ (2005). Antimicrobial activity of flavonoids. *Int. J. Antimicrobial Agents*, **26**: 343-356.
- Dao TT, Chi YS, Kim J, Kim HP, Kim S and Park H (2004). Synthesis and inhibitory activity against COX-2 catalyzed prostaglandin production of chrysin derivatives. *Bioorg. Med. Chem. Lett.*, **14**: 1165-1167.
- Fan Z, Zhao W, Guo J, Cheng R, Zhao J, Yang W, Wang Y, Li W and Peng X (2012). Antidepressant activities of flavonoids from *Glycyrrhiza uralensis* and its neurogenesis protective effect in rats. *Acta. Pharmaceutica. Sinica.*, **47**: 1612-1617.
- Fernández SP, Wasowski C, Loscalzo LM, Granger RE, Johnston GA, Paladini AC and Marder M (2006). Central nervous system depressant action of flavonoid glycosides. *Eur. J. Pharmacol.*, **539**: 168-176.
- Gross M (2004). Flavonoids and cardiovascular disease. *Pharmaceutical Biology*, **42**: 21-35.
- Imam MZ and Sumi CD (2014). Evaluation of antinociceptive activity of hydromethanol extract of *Cyperus rotundus* in mice. *BMC Complement. Altern. Med.*, **14**: 83.
- Kim HP, Son KH, Chang HW and Kang SS (2004). Anti-inflammatory plant flavonoids and cellular action mechanisms. *J. Pharmacol. Sci.*, **96**: 229-245.
- Koster R, Anderson M and Debeer E (1959). Acetic acid for analgesic screening. Federation proceedings, Federation amer soc exp biol 9650 rockville pike, Bethesda, MD 20814-3998: 412-412.
- Kris-Etherton P, Lefevre M, Beecher G, Gross M, Keen C and Etherton T (2004). Bioactive compounds in nutrition and health-research methodologies for establishing biological function: The antioxidant and anti-inflammatory effects of flavonoids on atherosclerosis. *Annu. Rev. Nutr.*, **24**: 511-538.
- Lorke D (1983). A new approach to practical acute toxicity testing. *Archive. Toxicol.*, **54**: 275-287.
- Maikai V (2011). Antitrypanosomal Activity of Flavonoid Extracted from *Ximenia Americana* Stem Bark. *Int. J. Biol.*, **3**.
- Middleton JRE and Kandaswami C (1992). Effects of flavonoids on immune and inflammatory cell functions. *Biochem. Pharmacol.*, **43**: 1167-1179.
- Moon YJ, Wang X and Morris ME (2006). Dietary flavonoids: Effects on xenobiotic and carcinogen metabolism. *Toxicol. in vitro.*, **20**: 187-210.
- Mostahar S, Alam S and Islam A (2006). Cytotoxic and antimicrobial activities of some synthetic flavones. *Indian J. Chem. Sec B.*, **45**: 1478.
- Mustaffa F, Indurkar J, Ismail S, Mordi M, Ramanathan S and Mansor S (2010). Analgesic activity, toxicity study and phytochemical screening of standardized *Cinnomomum iners* leaves methanolic extract. *Pharmacognosy Res.*, **2**: 76.
- Peterson J and Dwyer J (1998). Flavonoids: dietary occurrence and biochemical activity. *Nutrition Research*, **18**: 1995-2018.
- Ray D, Sharatchandra K and Thokchom I (2006). Antipyretic, antidiarrhoeal, hypoglycaemic and hepatoprotective activities of ethyl acetate extract of *Acacia catechu* Willd. in albino rats. *Indian J. Pharmacol.*, **38**.
- Razmi A, Zarghi A, Arfaee S, Naderi N and Faizi M (2013). Evaluation of Anti-nociceptive and Anti-inflammatory Activities of Novel Chalcone Derivatives. *IJPR*, **12**: 153.
- Rice-Evans CA, Miller NJ and Paganga G (1996). Structure-antioxidant activity relationships of flavonoids and phenolic acids. *Free Radic. Biol. Med.*, **20**: 933-956.
- Sandhar HK, Kumar B, Prasher S, Tiwari P, Salhan M and Sharma P (2011). A review of phytochemistry and pharmacology of flavonoids. *Int. Pharmaceutica. Sci.*, **1**: 25-41.
- Shah S and Goswami K (2013). Synthesis, characterization and anti microbial activity of some novel chalcone compounds having benzyloxy-monochloro resacetophenone moiety. *Der. Pharma. Chemica.*, **5**: 75-80.
- Simmons DL, Botting RM and Hla T (2004). Cyclooxygenase isozymes: The biology of prostaglandin synthesis and inhibition. *Pharmacol. Rev.*, **56**: 387-437.
- Smith PC, Thomas P, Scurr J and Dormandy J (1988). Causes of venous ulceration: A new hypothesis. *Br. Med. J. (Clinical research ed.)*, **296**: 1726.
- Srinivasan K, Muruganandan S, Lal J, Chandra S, Tandan SK, Raviprakash V and Kumar D (2003). Antinociceptive and antipyretic activities of *Pongamia pinnata* leaves. *Phytother. Res.*, **17**: 259-264.
- Subhan N, Alam A, Ahmed F and Shahid IZ (2008). Antinociceptive and gastroprotective effect of the crude ethanolic extracts of *Excoecaria agallocha* Linn. *Turk. J. Pharm. Sci.*, **5**: 143-154.
- Tijburg L, Mattern T, Folts J, Weisgerber U and Katan M (1997). Tea flavonoids and cardiovascular diseases: a review. *Critic. Rev. Food Sci. Nutrition.*, **37**: 771-785.
- Tripathi K (2013). *Essentials of Medical Pharmacology*, JP Medical Ltd.
- Wightman J Red berries and their health benefits. ACS symposium series, 2004. ACS Publications, pp.123-132.
- Woolfe G and MacDonald A (1944). The evaluation of the analgesic action of pethidine hydrochloride (Demerol). *J. Pharmacol. Experiment. Therap.*, **80**: 300-307.
- Zhang HY, Yang DP and Tang GY (2006). Multipotent antioxidants: From screening to design. *Drug. discovery. Today*, **11**: 749-754.